# Inference of biological networks: from de novo to supervised approaches 

Jean-Philippe Vert

Jean-Philippe.Vert@mines.org

Mines ParisTech / Institut Curie / Inserm
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## Outline

(9) Introduction
(2) De novo network inference

3 Supervised network inference: local models
(4) Supervised network inference: global models
(5) From local models to pairwise kernels
(6) Conclusion

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## Gene expression



Image adapted from: National Human Genome Research Institute.

## Gene expression regulation



## Gene regulatory network



## Gene regulatory network of E. coli



## Gene expression data



## Reconstruction of gene regulatory network from expression data



## More networks...



## More data..



- Gene expression measurements
- Phylogenetic profiles
- Location of proteins/enzymes in the cell
- Structures...


## General gene network inference problem



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## De novo inference

## The problem

Given data about the genes (eg, expression), infer the edges (eg, regulations).


## How?

- Interactions are between "similar" genes?
- Interactions are between "dependent" genes?
- Interactions are between "predictive" genes?


## De novo inference

## The problem

Given data about the genes (eg, expression), infer the edges (eg, regulations).


## How?

- Interactions are between "similar" genes?
- Interactions are between "dependent" genes?
- Interactions are between "predictive" genes?


## Predict interactions between "similar" genes

- In most networks, connected genes are significantly more "similar" than non-connected ones
- Inference: connect genes whose similarity (eg, Euclidean distance between profiles) is above a threshold


## Example: yeast metabolic network

- 769 proteins, 3702 metabolic edges
- Inference: rank by decreasing similariy of expression, interactions, localization, phylogenetic profiles



## Example: E coli regulatory network

- 154 TF targeting 1164 genes through 3293 regulations
- Inference: rank by decreasing Euclidean distance between expression profiles



## Predict regulations between "dependent" genes

Sometimes the expression of a TF and its target are not similar, but correlated or dependent



We can therefore try to detect these dependencies to infer regulation.

## Measuring dependency

## Pearson/Spearman correlation, mutual information (ARACNE, CLR...)



## Validation

## Application: E coli regulatory network : 154 TF targeting 1164 genes through 3293 regulations




## Predict regulations between "predictive" genes

- The dynamic equation of the mRNA concentration of a gene is of the form:

$$
\frac{d X}{d t}=f(X, R)
$$

where $R$ represent the set of concentrations of transcription factors that regulate $X$.

- At steady state, $d X / d t=0=f(X, R)$
- If we linearize $f(X, R)=0$ we get linear relation of the form

$$
X=\sum_{i \in R} \beta_{i} X_{i}
$$

- This suggests to look for transcription factors whose expression is sufficient to explain the expression of $X$ across different experiments.


## Predicting regulation by sparse regression

- Treat each target in turn
- Let $Y$ the expression of a target, and $X_{1}, \ldots, X_{p}$ the expression of all TFs. We look for a model

$$
Y=\sum_{i=1}^{p} \beta_{i} X_{i}+\text { noise }
$$

where $\beta$ is sparse, i.e., only a few $\beta_{i}$ are non-zero

- Examples:
- GENIE: feature selection by random forest (Huynh-Thu et al., 2010)
- Feature selection by Lasso + stability selection (Haury et al., 2011)
- Both methods were ranked 1st and 2nd (out of 28) at the DREAM5 in silico network inference challenge


## Summary on de novo network inference

- Feature selection methods seem to be state-of-the-art
- Performance remains low: recall below $10 \%$ for the best-known network
- How to infer the $90 \%$ of difficult interactions??
- improve de novo methods
- change the paradigm


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## Motivations



- In many cases, we already know quite a few regulations.
- Can we use them, in addition to expression data, to predict unknown regulations?


## Change of paradigm



- New hypothesis: genes regulated by the same TF have similar expression variations
- Note that this is very different from de novo inference, where we compare the expression profile of the gene to that of the TF
- Caveats:
- We need known interactions
- We may not find completely different interactions from those we know


## One-class learning approaches

- For a given TF, let $P \subset[1, n]$ be the set of genes known to be regulated by it
> - From the expression profiles $\left(X_{i}\right)_{i \in P}$, estimate a score $s(X)$ to assess which expression profiles $X$ are similar - Then classify the genes not in $P$ by decreasing score



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## Estimating the scoring function: examples



- Kernel density estimation

$$
s(X)=\sum_{i \in P} \exp \left(-\gamma\left\|X-X_{i}\right\|^{2}\right)
$$

- One-class SVM

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## From one-class to PU learning



- One class: given genes in $P$, estimate the function $s(X)$
- PU learning: given genes in $P$ and the set of unlabeled genes $U$, estimate the scores $s\left(X_{j}\right)$ for $j \in U$


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## Why PU learning can be better than one-class learning



## PU learning in practice


(1) Train a classifier to discriminate $P$ from $U$ (eg, SVM or random forest)
(2) Rank genes in $U$ by decreasing training score
(3) Bagging PU discrimination can help (Mordelet and V., 2010)

## One-class vs PU learning



More in Fantine Mordelet's PhD (2010)

## Supervised vs de novo inference




| Method | Recall at 60\% | Recall at 80\% |
| :--- | :---: | :---: |
| SIRENE | $\mathbf{4 4 . 5 \%}$ | $\mathbf{1 7 . 6 \%}$ |
| CLR | $7.5 \%$ | $5.5 \%$ |
| Relevance networks | $4.7 \%$ | $3.3 \%$ |
| ARACNe | $1 \%$ | $0 \%$ |
| Bayesian network | $1 \%$ | $0 \%$ |

SIRENE = Supervised Inference of REgulatory NEtworks (Mordelet and V., 2008)

## Application: predicted regulatory network (E. coli)



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## Motivations



- Local models require enough known targets of each TF. Can we share information across TF?
- For undirected networks (eg, PPI), how to reconcile local predictions?
- Idea: work directly in the space of pairs, to discriminate interacting vs non-interacting pairs.


## Pattern recognition for pairs: basic issue

- A pair can be connected (1) or not connected (-1)
- From the known subgraph we can extract examples of connected and non-connected pairs
- However the genomic data characterize individual proteins; we need to work with pairs of proteins instead!



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Known graph


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## Direct sum for ordered pairs?

- Each individual protein is represented by a vector $v \in \mathbb{R}^{p}$
- How to represent a pair of proteins $(u, v)$ by a vector $\psi(u, v) \in \mathbb{R}^{q}$ ?
- A simple idea is to concatenate the vectors $u$ and $v$ to obtain a $2 p$-dimensional vector of $(u, v)$ :
- Problem: a linear function then becomes additive...

$$
f(u, v)=w^{\top} \psi(u, v)=w_{1}^{\top} u+w^{\top} v
$$

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## Direct product for ordered pairs

- Alternatively, make the direct product, i.e., the $p^{2}$-dimensional vector whose entries are all products of entries of $u$ by entries of $v$ :

$$
\psi(u, v)=u \otimes v
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- Good news: inner product factorizes:

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$$
\left(u_{1} \otimes v_{1}\right)^{\top}\left(u_{2} \otimes v_{2}\right)=\left(u_{1}^{\top} u_{2}\right) \times\left(v_{1}^{\top} v_{2}\right)
$$

which is good for algorithms that use only inner products (SVM...):

$$
K_{P}\left(\left(u_{1}, v_{1}\right),\left(u_{2}, v_{2}\right)\right)=\psi\left(u_{1}, v_{1}\right)^{\top} \psi\left(u_{2}, v_{2}\right)=K\left(u_{1}, u_{2}\right) K\left(v_{1}, v_{2}\right)
$$

## Representing an unordered pair: TPPK

- Often we want to work with unordered pairs, e.g., PPI network:

$$
\{u, v\}=\{(u, v),(v, u)\}
$$

- This suggest to symmetrize the representation of ordered pairs:

- This leads to the symmetric tensor product pairwise kernel (TPPK) (Ben-Hur and Noble, 2006):


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$K_{\text {TPPK }}\left(\left\{u_{1}, v_{1}\right\},\left\{u_{2}, v_{2}\right\}\right)=K\left(u_{1}, u_{2}\right) K\left(v_{1}, v_{2}\right)+K\left(u_{1}, v_{2}\right) K\left(v_{1}, u_{2}\right)$


## Another representation: MLPK

- Another symmetric representation:

$$
\psi(\{u, v\})=(u-v)^{\otimes 2}
$$

- Equivalently, train the SVM over pairs with the metric learning pairwise kernel:

$$
\begin{aligned}
& K_{M L P K}\left(\left\{u_{1}, v_{1}\right\},\left\{u_{2}, v_{2}\right\}\right)=\psi\left(\left\{u_{1}, v_{1}\right\}\right)^{\top} \psi\left(\left\{u_{2}, v_{2}\right\}\right) \\
& \quad=\left[K\left(u_{1}, u_{2}\right)-K\left(u_{1}, v_{2}\right)-K\left(v_{1}, u_{2}\right)+K\left(v_{1}, v_{2}\right)\right]^{2}
\end{aligned}
$$

- Theorem: A SVM with the MLPK kernel trained to discriminate connected from non-connected pairs, solves a metric learning problem (V. et al., 2007)


## Technical details

- For two vectors $u, v \in \mathcal{H}$ let the metric:

$$
d_{M}(u, v)=(u-v)^{\top} M(u-v)
$$

- Learn the metric so that points close to each other are connected?
- We consider the problem:

where I is a hinge loss to enforce:

- SVM with MLPK kernel solve it without the constraint $M \geq 0$


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$$
\min _{M \geq 0} \sum_{i} I\left(u_{i}, v_{i}, y_{i}\right)+\lambda\|M\|_{\text {Frobenius }}^{2}
$$

where I is a hinge loss to enforce:

$$
d_{M}\left(u_{i}, v_{i}\right) \begin{cases}\leq 1-\gamma & \text { if }\left(u_{i}, v_{i}\right) \text { is connected } \\ \geq 1+\gamma & \text { otherwise }\end{cases}
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## Alternative: symmetrized local models for undirected networks

The idea (Bleakley and V., 2007)

- For each protein $P$, make a local model using known partners as positive examples to estimate an interaction score $s_{P}\left(P^{\prime}\right)$ for any candidate partner $P^{\prime}$
- Symmetrize a posteriori: the interaction score of a candidate pair $P, P^{\prime}$ is:

$$
s_{P}\left(P^{\prime}\right)+s_{P^{\prime}}(P)
$$



## Results: protein-protein interaction (yeast)



(from Bleakley et al., 2007)

## Results: metabolic gene network (yeast)



(from Bleakley et al., 2007)

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## Motivation

In the case of unordered pairs $\{A, B\}$, pairwise kernels such as the TPPK and local models look very different:

- Local models seem to over-emphasize the asymmetry of the relationships, but symmetrize the prediction a posteriori
- Pairwise kernels symmetrize the data a priori and learn in the space or unordered pairs
Can be clarify the links between these approaches, and perhaps interpolate between them?


## Notations

- $\mathcal{A}$ the set of individual proteins, endowed with a kernel $K_{\mathcal{A}}$
- $\mathcal{X}=\mathcal{A}^{2}$ the set of ordered pairs of the form $x=(a, b)$ endowed with a kernel $K_{\mathcal{X}}$ (usually deduced from $K_{\mathcal{A}}$ )
- $\mathcal{P}$ the set of unordered pairs of the form $p=\{(a, b),(b, a)\}$
- We want to learn over $\mathcal{P}$ from a set of labeled training pairs $\left(p_{1}, y_{1}\right), \ldots,\left(p_{n}, y_{n}\right) \in \mathcal{P} \times\{-1,1\}$



## Two strategies to learn over $\mathcal{P}$

## Strategy 1: Inference over $\mathcal{P}$ with a pair kernel

(1) Define a kernel $K_{\mathcal{P}}$ over $\mathcal{P}$ by convolution of $K_{\mathcal{X}}$ :

$$
K_{\mathcal{P}}\left(p, p^{\prime}\right)=\frac{1}{|p| \cdot\left|p^{\prime}\right|} \sum_{x \in p, x^{\prime} \in p^{\prime}} K_{\mathcal{X}}\left(x, x^{\prime}\right) .
$$

(2) Train a classifier over $\mathcal{P}$ e.g., a SVM, using the kernel $K_{\mathcal{P}}$

## Strategy 2: Inference over $\gamma$ with a pair duplication <br> (1) Duplicate each training pair $p=\{a, b\}$ into 2 ordered paired <br> (2) Train a classifier over $\mathcal{X}$, e.g., a SVM, using the kernel $K_{\mathcal{X}}$ <br> (3) The classifier over $\mathcal{P}$ is then the a posteriori average:

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$$
f_{\mathcal{P}}(p)=\frac{1}{|p|} \sum_{x \in p} f_{\mathcal{X}}(x)
$$

## The TPPK kernel

$$
K_{T P P K}(\{a, b\},\{c, d\})=K_{\mathcal{A}}(a, c) K_{\mathcal{A}}(b, d)+K_{\mathcal{A}}(a, d) K_{\mathcal{A}}(b, c) .
$$

## Theorem

Let $\mathcal{X}=\mathcal{A}^{2}$ be endowed with the p.d. kernel:

$$
\begin{equation*}
K_{\mathcal{X}}((a, b),(c, d))=2 K_{\mathcal{A}}(a, c) K_{\mathcal{A}}(b, d) \tag{1}
\end{equation*}
$$

Then the TPPK approach is equivalent to both Strategy 1 and Strategy 2.

Remarks: Equivalence with Strategy 1 is obvious, equivalence with Strategy 2 is not, see proof in Hue and V. (ICML 2010).

## The local models



## Theorem

Let $\mathcal{X}=\mathcal{A}^{2}$ be endowed with the p.d. kernel:

$$
K_{\mathcal{X}}((a, b),(c, d))=\delta(a, c) K_{\mathcal{A}}(b, d)
$$

where $\delta$ is the Kronecker kernel $(\delta(a, c)=1$ if $a=c, 0$ otherwise). Then the local approach is equivalent to Strategy 2.

Remarks: Strategies 1 and 2 are not equivalent with this kernel. In general, they are equivalent up to a modification in the loss function of the learning algorithm, see details in Hue and V. (ICML 2010)..

## Interpolation between local model and TPPK

|  | Strategy 1: pair kernel | Strategy 2: duplication |
| :---: | :---: | :---: |
| $K_{\mathcal{X}}=K_{\mathcal{A}} \otimes K_{\mathcal{A}}$ | TPPK | TPPK |
| $K_{\mathcal{X}}=\delta \otimes K_{\mathcal{A}}$ | new | Local model |

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Interpolation:

$$
K_{\mathcal{X}}=\left((1-\lambda) K_{\mathcal{A}}+\lambda \delta\right) \otimes K_{\mathcal{A}}
$$

for $\lambda \in[0,1]$

## Interpolation kernel




Metabolic networks with localization data (left); PPI network with expression data (right)

## Interpolation kernel

Table: Strategy and kernel realizing the maximum mean AUC for nine metabolic and protein-protein interaction networks experiments, with the kernel $K^{\lambda}$ for $\lambda \in[0,1]$.

| benchmark | best kernel |
| :---: | :---: |
| interaction, exp | Duplicate, $\lambda=0.7$ |
| interaction, loc | Pair kernel, $\lambda=0.6$ |
| interaction, phy | Duplicate, $\lambda=0.8$ |
| interaction, y2h | Duplicate / Pair kernel, $\lambda=0$ |
| interaction, integrated | Duplicate / Pair kernel, $\lambda=0$ |
| metabolic, exp | Pair kernel, $\lambda=0.6$ |
| metabolic, loc | Pair kernel, $\lambda=1$ |
| metabolic, phy | Pair kernel, $\lambda=0.6$ |
| metabolic, integrated | Duplicate / Pair kernel, $\lambda=0$ |

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Take-home messages

- De novo inference: feature selection methods state-of-the-art, but overall performance very limited (recall < 10\%)
- Supervised inference: the change of paradigm boosts the performance. Difficult to do better than local models.
- If you already know edges, supervised inference is much more powerful than de novo inference
- New ideas for de novo inference?
- More direct formulation as structured output learning?
- Better adjust the complexity of models to the complexity of the task?
- Link de novo and supervised inference?
- Combine edge inference with graph models? Link with methods in relational learning and collaborative filtering?


## Interpolation between local model and TPPK

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## Some interesting questions

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## Thanks!



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