

Some contributions of machine learning to bioinformatics

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Where I come from



Inserm

- A joint lab about “Cancer computational genomics, bioinformatics, biostatistics and epidemiology”
- Located in th Institut Curie, a major hospital and cancer research institute in Europe
- Hosted in a brand new building in the center of Paris (near Notre-Dame, Saint-Germain, Pantheon...)

Main topics

- **Epidemiology** of cancer (eg, studies on etiology of breast cancer)
- **General biostatistics** (eg, clinical trials, risk modelling...)
- **Biostatistics and machine learning for bioinformatics**
(high-throughput data processing, modeling, predictive models...)
- **Systems biology**: analysis, modeling, inference of important regulatory and signaling systems
- **IT**: software developments, DB, web

Main topics in machine learning / statistics

- Processing **high-throughput data** (normalization, analysis): transcriptome, genome (CGH/SNP), proteomics, kinome. High-throughput sequencing is coming soon.
- Making **predictive models**, in particular diagnosis / prognosis
- **Data mining** / integration of **heterogeneous** data
- **Structural bioinformatics**, protein-protein interactions, virtual screening, chemogenomics

Outline

- 1 Including prior knowledge in classification and regression
- 2 Virtual screening and chemogenomics
- 3 Inference on biological networks
- 4 Conclusion

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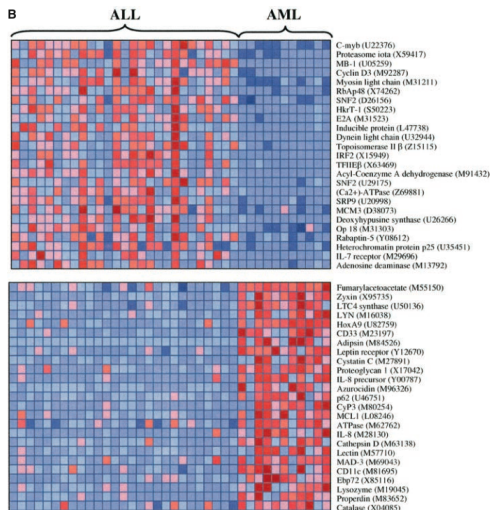
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Motivation



Goal

- Design a **classifier** to automatically assign a class to future samples from their expression profile
- **Interpret** biologically the differences between the classes

Difficulty

- Large dimension
- Few samples

The model

- Each sample is represented by a vector $x = (x_1, \dots, x_p)$
- **Goal**: estimate a linear function:

$$f_{\beta}(x) = \sum_{i=1}^p \beta_i x_i + \beta_0 .$$

- **Interpretability**: the weight β_i quantifies the influence of feature i (but...)

Training the model

$$f_{\beta}(x) = \sum_{i=1}^p \beta_i x_i + \beta_0 .$$

- Minimize an **empirical risk** on the training samples:

$$\min_{\beta \in \mathbb{R}^{p+1}} R_{emp}(\beta) = \frac{1}{n} \sum_{i=1}^n l(f_{\beta}(x_i), y_i) ,$$

- ... subject to some **constraint** on β , e.g.:

$$\Omega(\beta) \leq C .$$

Example : Norm Constraints

The approach

A common method in statistics to learn with few samples in high dimension is to **constrain the Euclidean norm of β**

$$\Omega_{ridge}(\beta) = \|\beta\|_2^2 = \sum_{i=1}^p \beta_i^2,$$

(ridge regression, support vector machines...)

Pros

- Good performance in classification

Cons

- Limited interpretation (small weights)
- No prior biological knowledge

Example : Feature Selection

The approach

Constrain most weights to be 0, i.e., **select a few genes** (< 100) whose expression are sufficient for classification.

- Greedy feature selection (T-tests, ...)
- Constrain the norm of β : **LASSO** penalty ($\|\beta\|_1 = \sum_{i=1}^p |\beta_i|$), **elastic net** penalty ($\|\beta\|_1 + \|\beta\|_2$), ...)

Pros

- Good performance in classification
- **Biomarker** selection
- Interpretability

Cons

- The gene selection process is usually **not robust**
- No use of prior biological knowledge

The idea

- If we have a specific **prior knowledge about the “correct” weights**, it can be included in Ω in the constraint:

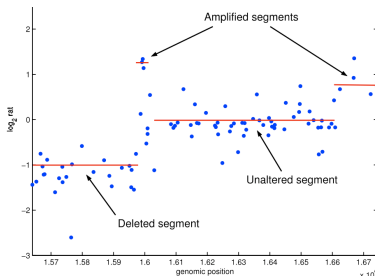
Minimize $R_{emp}(\beta)$ subject to $\Omega(\beta) \leq C$.

- If we design a **convex** function Ω , then the algorithm boils down to a convex optimization problem (usually **easy to solve**).
- Similar to priors in Bayesian statistics

Example: CGH array classification

The problem

- Comparative genomic hybridization (CGH) data measure the **DNA copy number** along the genome
- Very useful, in particular in cancer research
- Can we **classify CGH arrays** for diagnosis or prognosis purpose?
- **Prior knowledge**: we expect β to be **sparse**, and **piecewise constant** along the genome

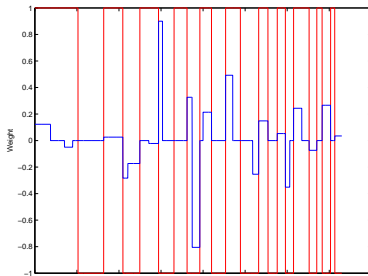
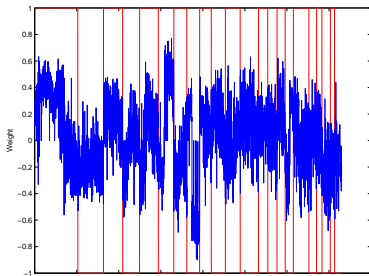


Example: CGH array classification

A solution (Rapaport et al., 2008)

$$\Omega_{fusedlasso}(\beta) = \sum_i |\beta_i| + \sum_{i \sim j} |\beta_i - \beta_j|.$$

- Good performance on diagnosis for bladder cancer, and prognosis for melanoma.
- More interpretable classifiers



Example: finding discriminant modules in gene networks

The problem

- Classification of gene expression: too many genes
- **A gene network is given** (PPI, metabolic, regulatory, signaling, co-expression...)
- We expect that “clusters of genes” (**modules**) in the network contribute similarly to the classification

Two solutions (Rapaport et al., 2007, 2008)

$$\Omega_{\text{spectral}}(\beta) = \sum_{i \sim j} (\beta_i - \beta_j)^2,$$

$$\Omega_{\text{graphfusion}}(\beta) = \sum_{i \sim j} |\beta_i - \beta_j| + \sum_i |\beta_i|.$$

Example: finding discriminant modules in gene networks

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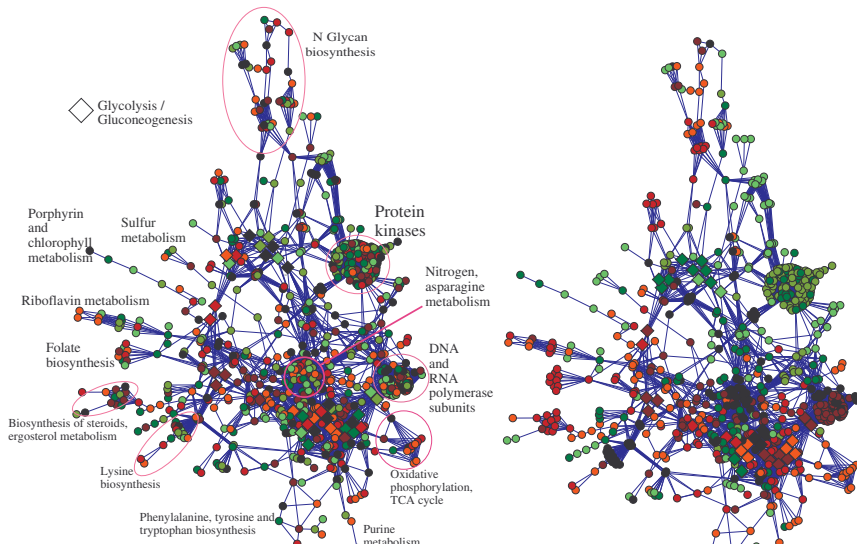
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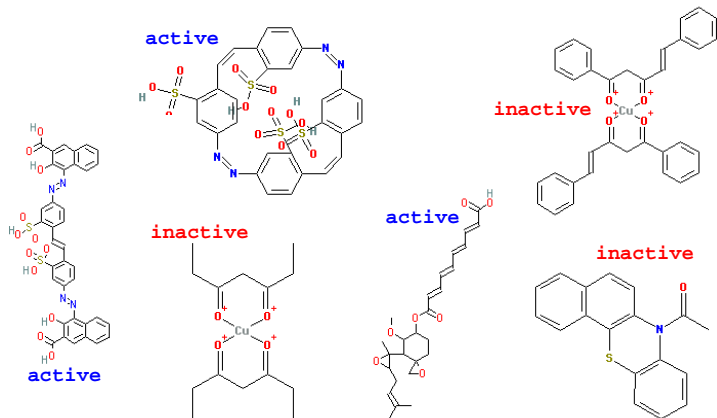
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Example: finding discriminant modules in gene networks



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Ligand-Based Virtual Screening and QSAR



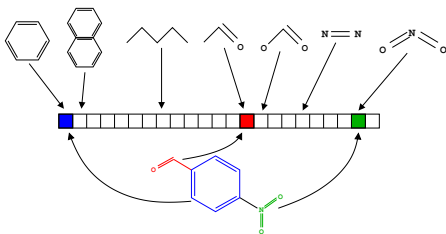
NCI AIDS screen results (from <http://cactus.nci.nih.gov>).

Classical approaches

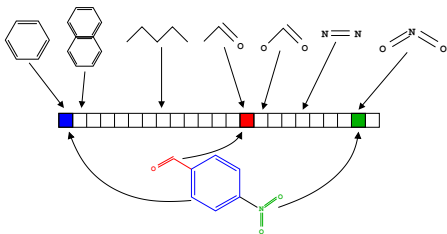
Two steps

- 1 Map each molecule to a **vector of fixed dimension** using **molecular descriptors**
 - Global properties of the molecules (mass, logP...)
 - 2D and 3D descriptors (substructures, fragments,)
- 2 Apply an algorithm for **regression or pattern recognition**.
 - PLS, ANN, ...

Example: 2D structural keys



Which descriptors?



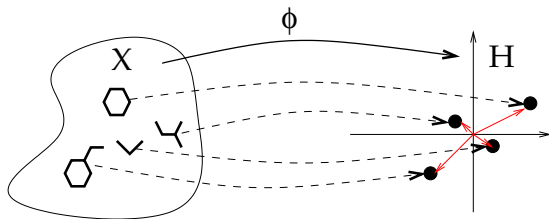
Difficulties

- **Many** descriptors are **needed** to characterize various features (in particular for 2D and 3D descriptors)
- But **too many** descriptors are **harmful** for memory storage, computation speed, statistical estimation

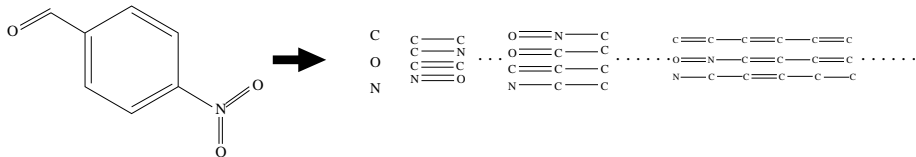
Definition

- Let $\Phi(x) = (\Phi_1(x), \dots, \Phi_p(x))$ be a vector representation of the molecule x
- The **kernel** between two molecules is defined by:

$$K(x, x') = \Phi(x)^\top \Phi(x') = \sum_{i=1}^p \Phi_i(x) \Phi_i(x').$$



Example: 2D fragment kernel



- $\phi_d(x)$ is the vector of counts of **all fragments of length d** :

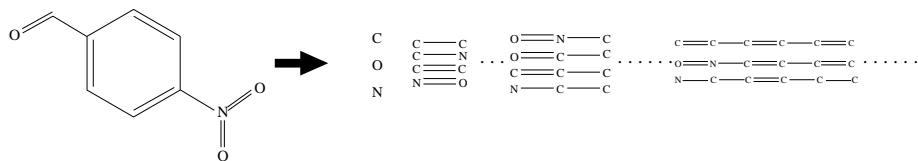
$$\phi_1(x) = (\#(C), \#(O), \#(N), \dots)^T$$

$$\phi_2(x) = (\#(C-C), \#(C=O), \#(C-N), \dots)^T \text{ etc...}$$

- The **2D fragment kernel** is defined, for $\lambda < 1$, by

$$K_{\text{fragment}}(x, x') = \sum_{d=1}^{\infty} r(\lambda) \phi_d(x)^T \phi_d(x').$$

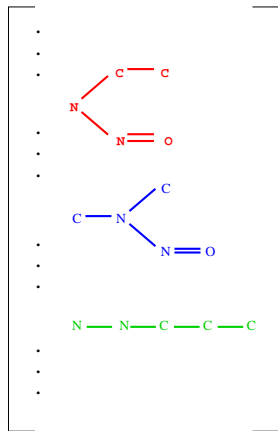
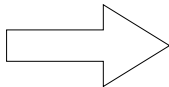
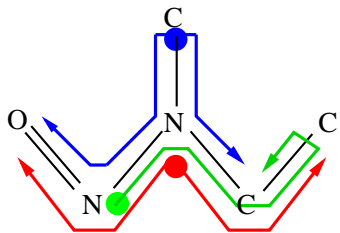
Example: 2D fragment kernel



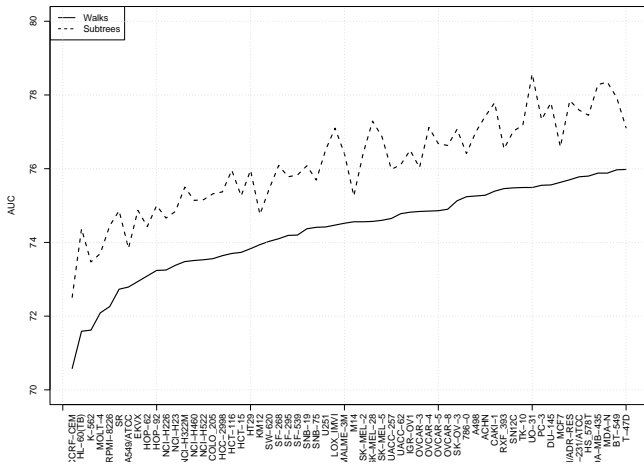
In practice

- $K_{fragment}$ can be **computed efficiently** (geometric kernel, random walk kernel...) although the feature space has **infinite dimension**.
- Increasing the **specificity of atom labels** improves performance
- Selecting only **“non-tottering” fragments** can be done efficiently and improves performance.

Example: 2D subtree kernel

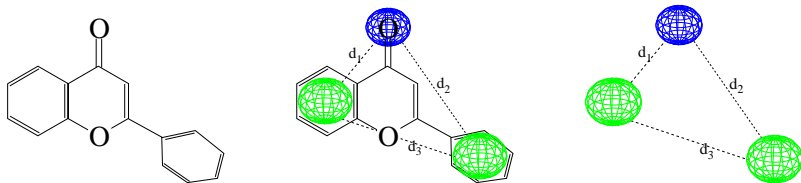


2D Subtree vs fragment kernels (Mahé and V, 2007)



Screening of inhibitors for 60 cancer cell lines (from Mahé and V., 2008)

Example: 3D pharmacophore kernel (Mahé et al., 2005)



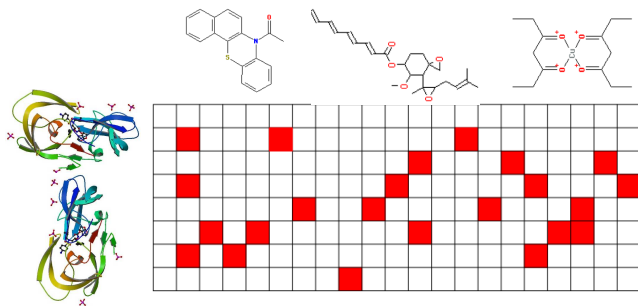
$$K(x, y) = \sum_{p_x \in \mathcal{P}(x)} \sum_{p_y \in \mathcal{P}(y)} \exp(-\gamma d(p_x, p_y)) .$$

Results (accuracy)

Kernel	BZR	COX	DHFR	ER
2D (Tanimoto)	71.2	63.0	76.9	77.1
3D fingerprint	75.4	67.0	76.9	78.6
3D not discretized	76.4	69.8	81.9	79.8

The problem

- Similar targets bind similar ligands
- Instead of focusing on each target individually, can we screen the biological space (target families) vs the chemical space (ligands)?
- Mathematically, learn $f(\text{target}, \text{ligand}) \in \{\text{bind}, \text{notbind}\}$



Tensor product SVM

- Take the kernel:

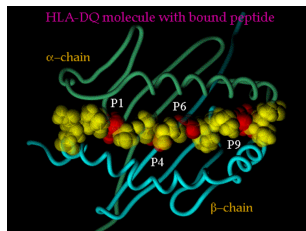
$$K((t, l), (t', l')) = K_t(t, t')K_l(l, l').$$

- Equivalently, represent a pair (t, l) by the vector $\phi_t(t) \otimes \phi_l(l)$
- Allows to use **any** kernel for proteins K_t with **any** kernel for small molecules K_l
- When K_t is the **Dirac** kernel, we recover the **classical paradigm**: each target is treated independently from the others.
- Otherwise, information is **shared across targets**. The more similar the targets, the more they share information.

Example: MHC-I epitope prediction across different alleles

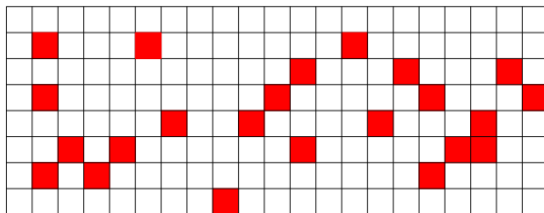
The approach (Jacob and V., 2007)

- take a kernel to compare different MHC-I alleles (e.g., based on the amino-acids in the peptide recognition pocket)
- take a kernel to compare different epitopes (9-mer peptides)
- Combine them to learn the $f(\text{allele}, \text{epitope})$ function
- State-of-the-art performance
- Available at <http://cbio.ensmp.fr/kiss>



Generalization: collaborative filtering with attributes

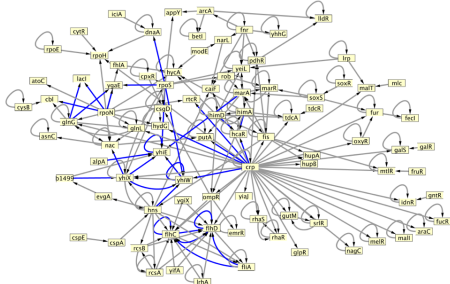
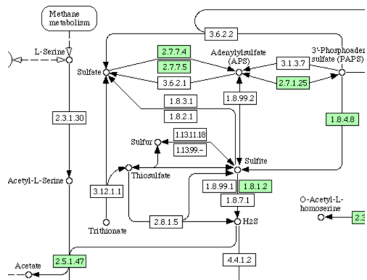
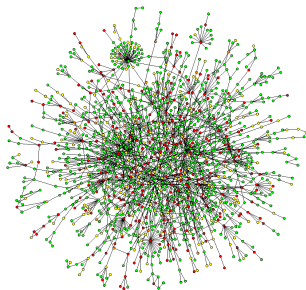
- General problem: learn $f(x, y)$ with a kernel K_x for x and a kernel K_y for y .
- SVM with a tensor product kernel $K_x \otimes K_y$ is a particular case of something more general: estimating an **operator** with a **spectral regularization**.
- Other spectral regularization are possible (e.g., **trace norm**) and lead to efficient algorithms
- More details in Abernethy et al. (2008).



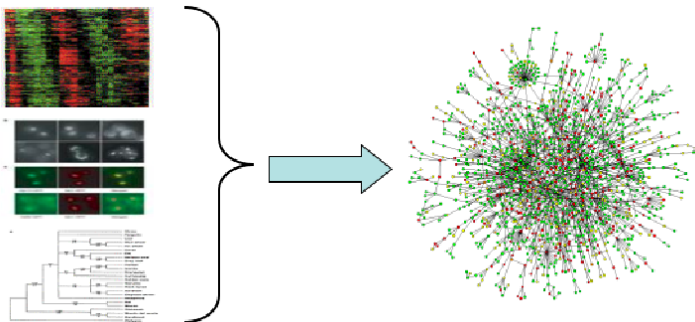
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Biological networks



Our goal



Data

- Gene expression,
- Gene sequence,
- Protein localization, ...

Graph

- Protein-protein interactions,
- Metabolic pathways,
- Signaling pathways, ...

“De novo” inference

- Given data about individual genes and proteins
- Infer the edges between genes and proteins

“Supervised” inference

- Given data about individual genes and proteins
- **and** given some known interactions
- infer unknown interactions

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Main messages

- 1 Most methods developed so far are “**de novo**” (e.g., co-expression, Bayesian networks, mutual information nets, dynamical systems...)
- 2 However most **real-world** application fit the “**supervised**” framework
- 3 Solving the “supervised” problem is **much easier** (and more efficient) than the “de novo” problem. It requires less hypothesis.

Typical strategies

- Fit a **dynamical system** to time series (e.g., PDE, boolean networks, state-space models)
- Detect **statistical conditional independence or dependency** (Bayesian network, mutual information networks, co-expression)

Pros

- **Excellent approach** if the model is correct and enough data are available
- **Interpretability** of the model
- Inclusion of **prior knowledge**

Cons

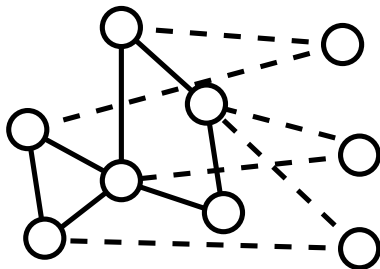
- **Specific** to particular data and networks
- **Needs a correct model!**
- Difficult **integration** of heterogeneous data
- Often needs a **lot of data** and long computation time

Supervised methods

Motivation

In actual applications,

- we know in advance parts of the network to be inferred
- the problem is to add/remove nodes and edges using genomic data as side information



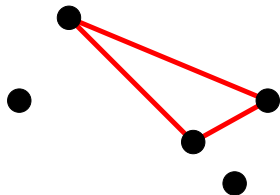
Supervised method

- Given genomic data **and** the currently known network...
- Infer **missing edges** between current nodes and additional nodes.

Supervised approach by Metric learning

Idea

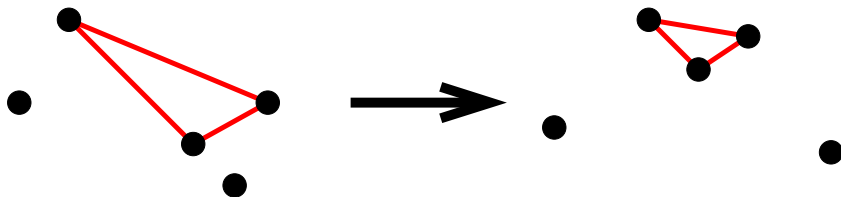
- The direct similarity-based method fails because the **distance metric used might not be adapted** to the inference of the targeted protein network.
- Solution: use the **known subnetwork** to **refine the distance measure**, before applying the similarity-based method
- Examples: **kernels CCA** (Yamanishi et al. 2004), **kernel metric learning** (V and Yamanishi, 2005)



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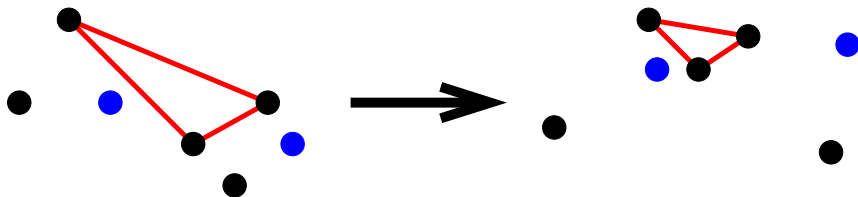
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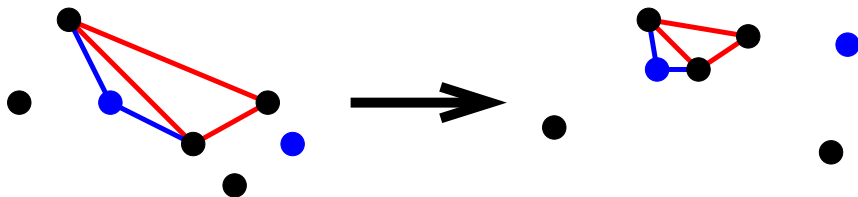
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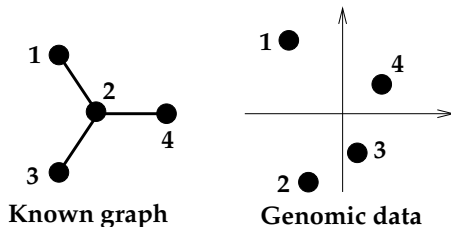
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Supervised inference by pattern recognition

Formulation and basic issue

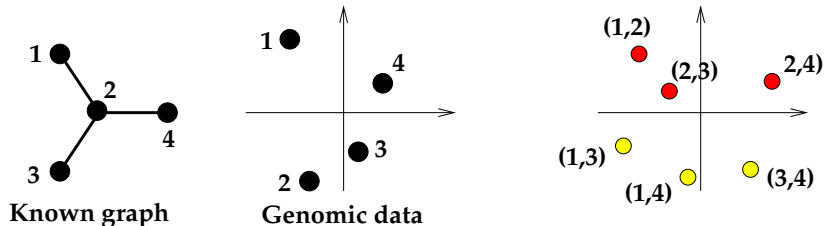
- A pair can be **connected (1)** or **not connected (-1)**
- From the known subgraph we can **extract examples** of connected and non-connected pairs
- However the genomic data characterize **individual** proteins; we need to work with **pairs** of proteins instead!



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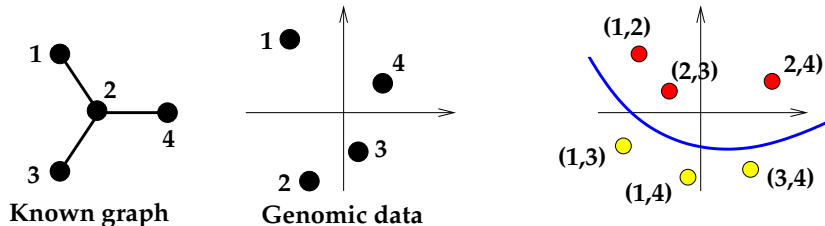
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Tensor product SVM (Ben-Hur and Noble, 2006)

- **Intuition:** a pair (A, B) is similar to a pair (C, D) if:
 - A is similar to C **and** B is similar to D , **or**...
 - A is similar to D **and** B is similar to C
- **Formally**, define a similarity between pairs from a similarity between individuals by

$$K_{TPPK}((a, b), (c, d)) = K(a, c)K(b, d) + K(a, d)K(b, c) .$$

- If K is a positive definite kernel for individuals then K_{TPPK} is a p.d. kernel for pairs which can be used by SVM
- This amounts to representing a pair (a, b) by the **symmetrized tensor product**:

$$(a, b) \rightarrow (a \otimes b) \oplus (b \otimes a) .$$

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Metric learning pairwise SVM (V. et al, 2007)

- **Intuition:** a pair (A, B) is similar to a pair (C, D) if:
 - $A - B$ is similar to $C - D$, or...
 - $A - B$ is similar to $D - C$.
- **Formally**, define a similarity between pairs from a similarity between individuals by

$$K_{MLPK}((a, b), (c, d)) = (K(a, c) + K(b, d) - K(a, d) - K(b, c))^2 .$$

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$$(a, b) \rightarrow (a - b)^{\otimes 2} .$$

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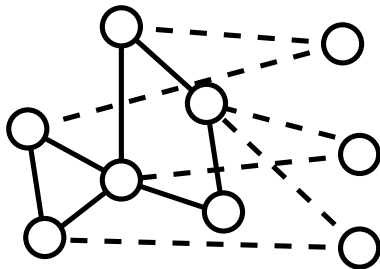
- If K is a positive definite kernel for individuals then K_{MLPK} is a p.d. kernel for pairs which can be used by SVM
- This amounts to representing a pair (a, b) by the **symmetrized difference**:

$$(a, b) \rightarrow (a - b)^{\otimes 2} .$$

Supervised inference with local models

The idea (Bleakley et al., 2007)

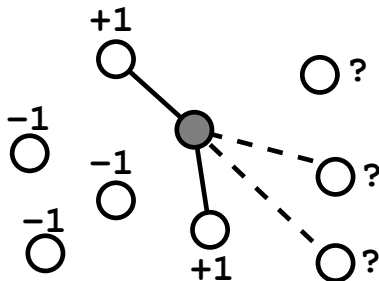
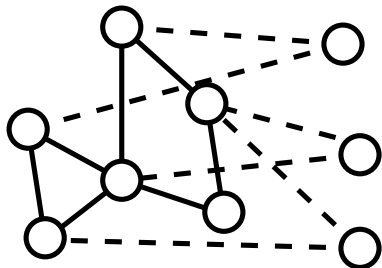
- Motivation: define **specific models** for **each target node** to discriminate between its neighbors and the others
- Treat each node independently from the other. Then **combine** predictions for ranking candidate edges.



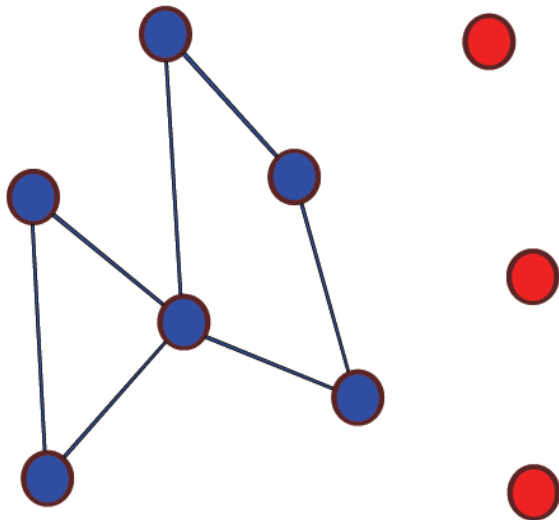
Supervised inference with local models

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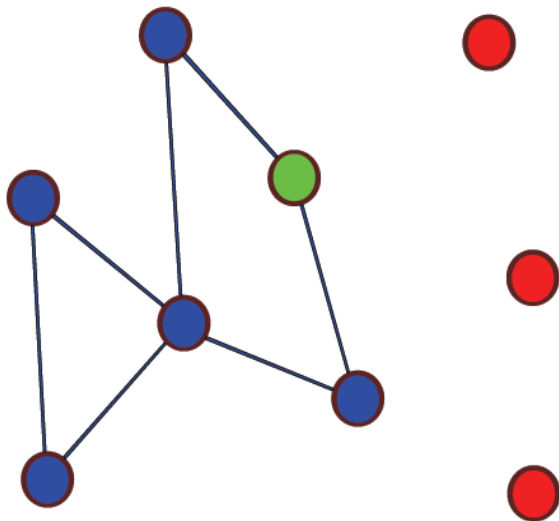
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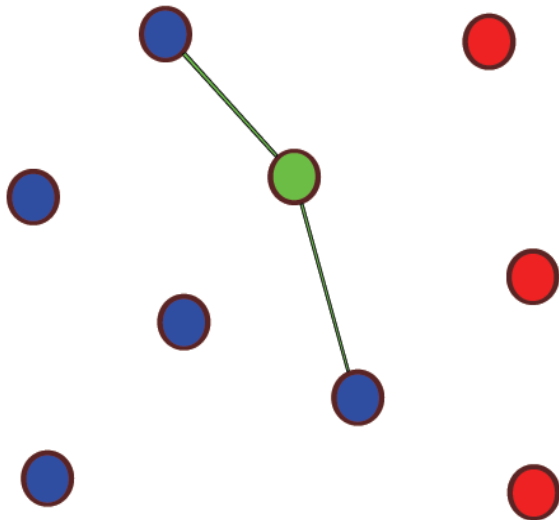
The LOCAL model



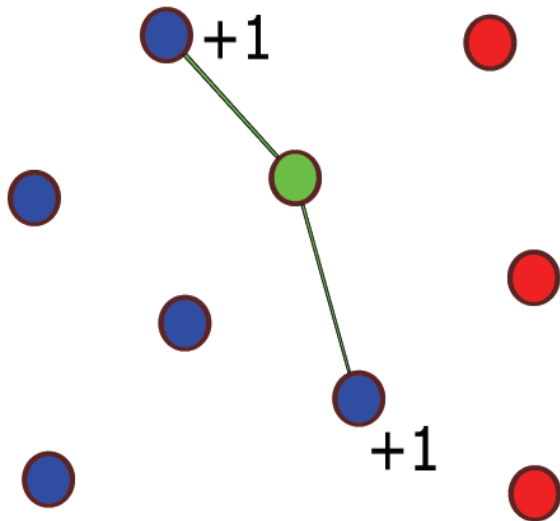
The LOCAL model



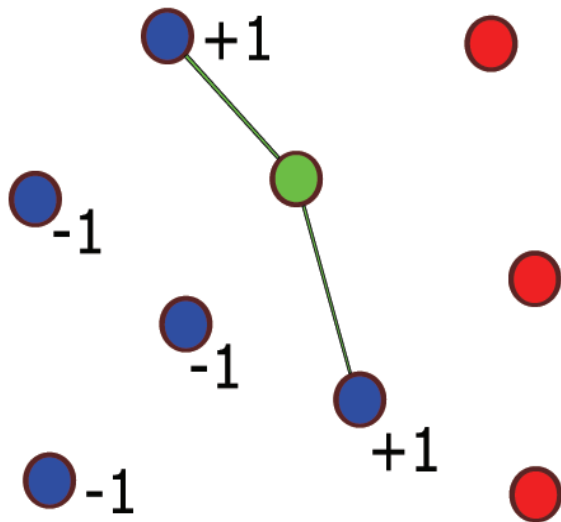
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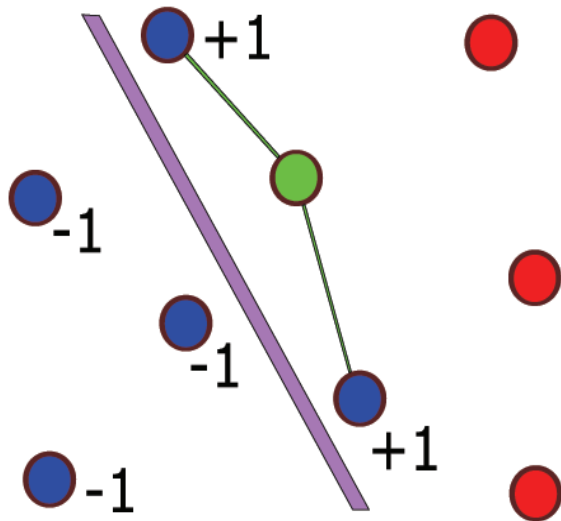
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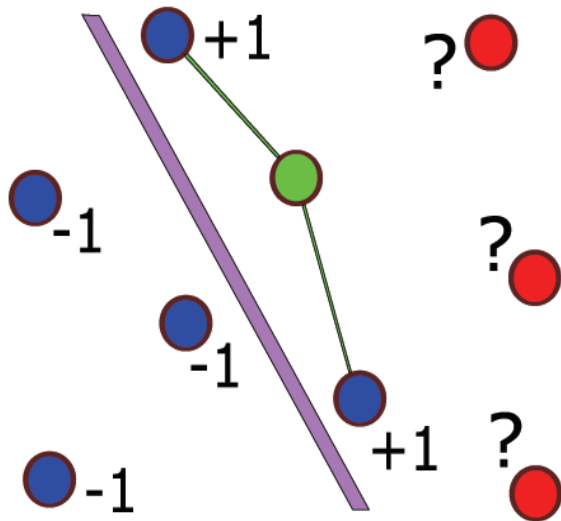
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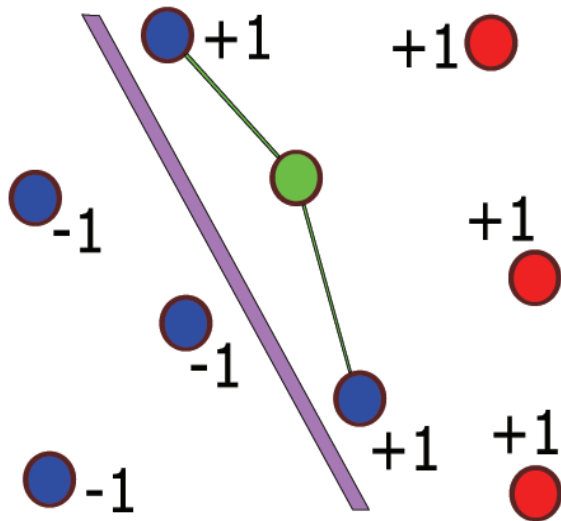
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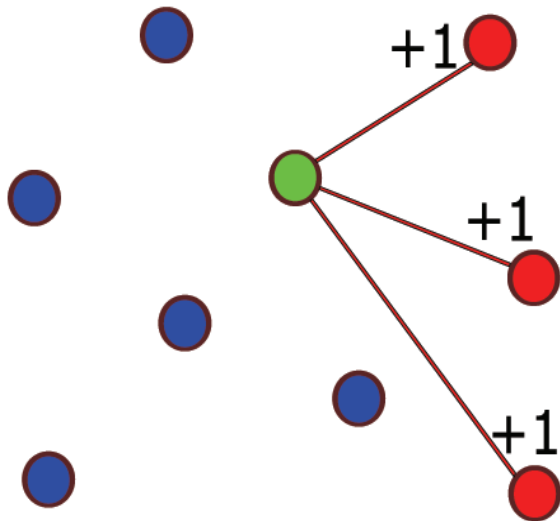
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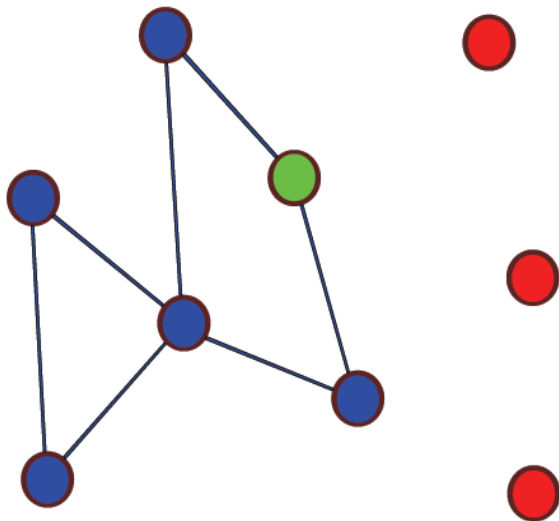
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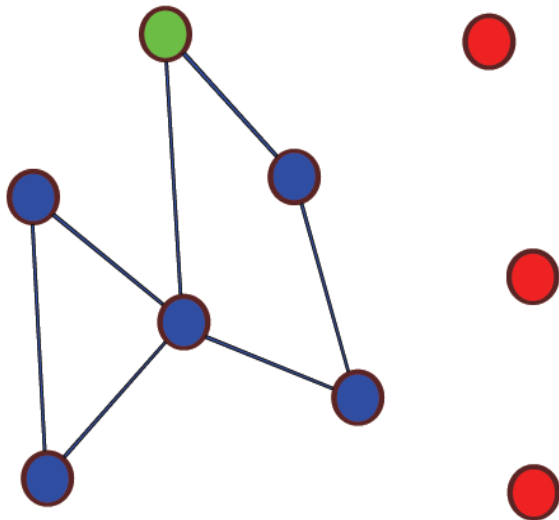
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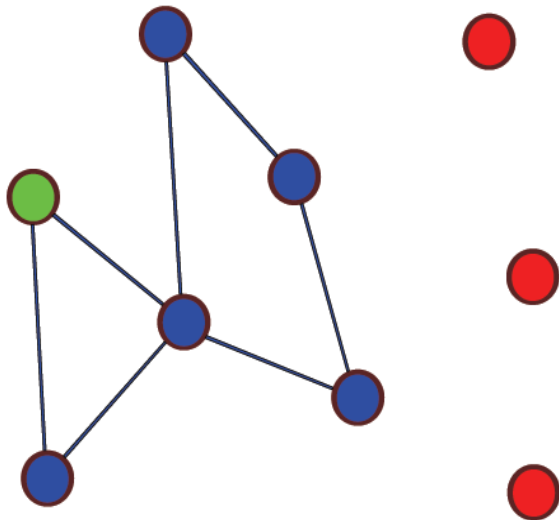
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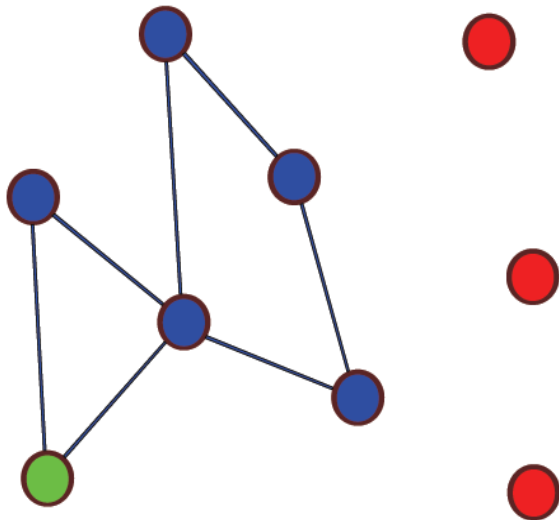
The LOCAL model



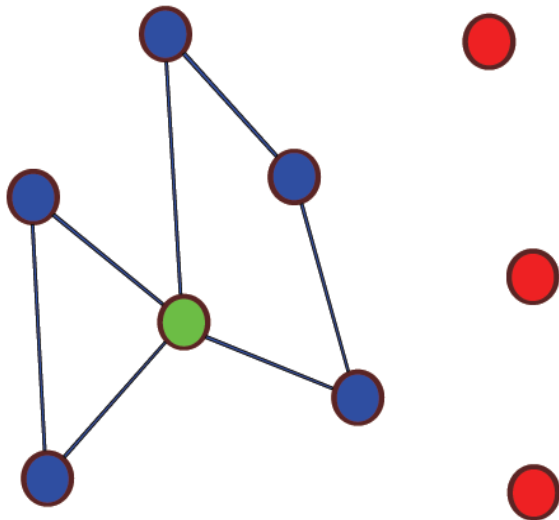
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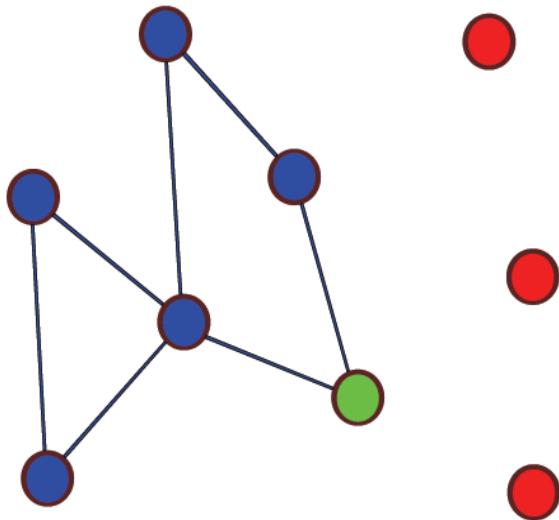
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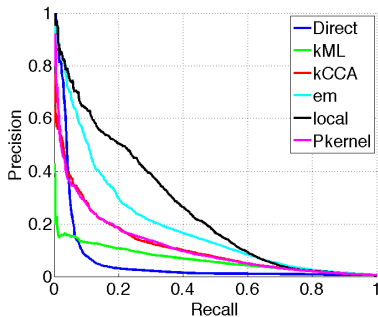
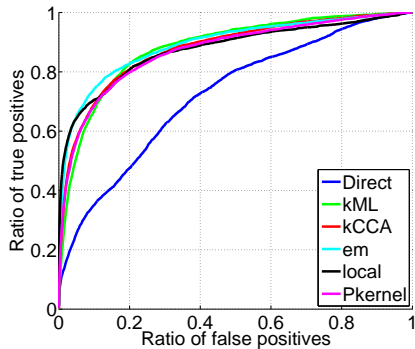
The LOCAL model



The LOCAL model

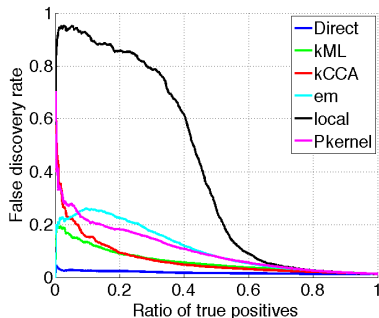
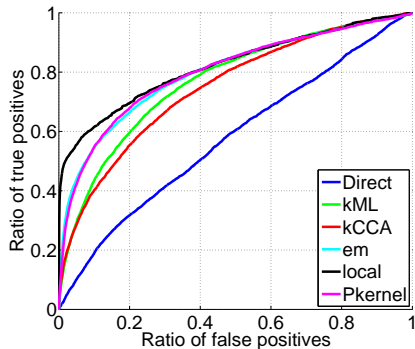


Results: protein-protein interaction (yeast)



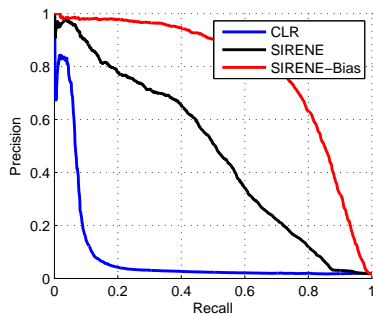
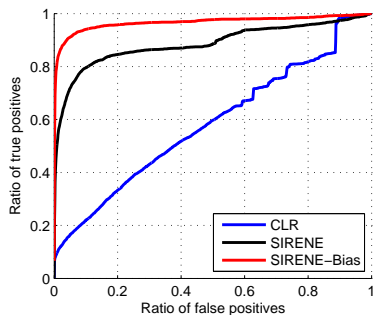
(from Bleakley et al., 2007)

Results: metabolic gene network (yeast)



(from Bleakley et al., 2007)

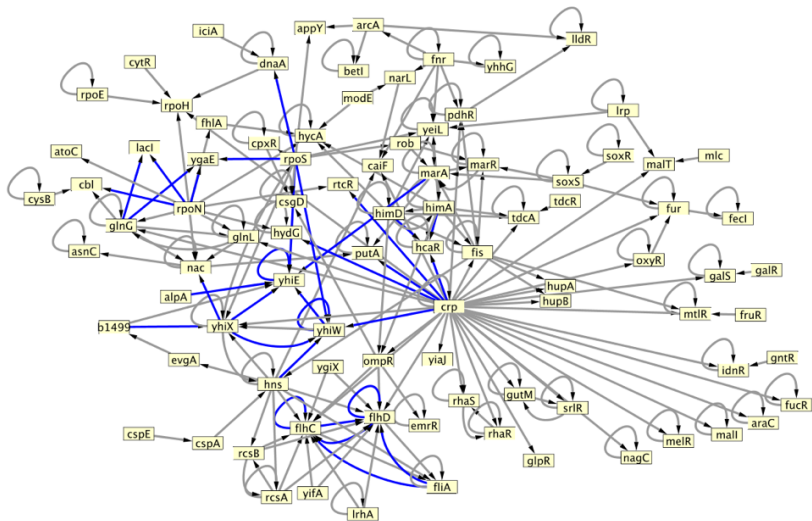
Results: regulatory network (E. coli)



Method	Recall at 60%	Recall at 80%
SIRENE	44.5%	17.6%
CLR	7.5%	5.5%
Relevance networks	4.7%	3.3%
ARACNe	1%	0%
Bayesian network	1%	0%

SIRENE = Supervised Inference of REgulatory Networks (Mordelet and V., 2008)

Results: predicted regulatory network (E. coli)



Prediction at 60% precision, restricted to transcription factors (from Mordelet and V., 2008).

Outline

- 1 Including prior knowledge in classification and regression
- 2 Virtual screening and chemogenomics
- 3 Inference on biological networks
- 4 Conclusion**

- Modern machine learning methods for regression / classification lend themselves well to the **integration of prior knowledge** in the penalization / regularization function, in particular for feature selection / grouping. Applications in **array CGH classification, siRNA design, microarray classification with gene networks**
- Kernel methods (eg SVM) allow to manipulate complex objects (eg molecules, biological sequences) as soon as **kernels can be defined and computed**. Applications in **virtual screening, QSAR, chemogenomics**.
- Inference of **biological networks** can be formulated as a **supervised problem** if the graph is partly known, and powerful methods can be applied. Application in **PPI, metabolic and regulatory networks inference**.

People I need to thank

Including prior knowledge in penalization

Franck Rapaport, Emmanuel Barillot, Andrei Zynoviev, Christian Lajaunie, Yves Vandenbrouck, Nicolas Foveau...

Virtual screening, kernels etc..

Pierre Mahé, Laurent Jacob, Liva Ralaivola, Véronique Stoven, Brice Hoffman, Martial Hue, Francis Bach, Jacob Abernethy, Theos Evgeniou...

Network inference

Kevin Bleakley, Fantine Mordelet, Yoshihiro Yamanihi, Gérard Biau, Minoru Kanehisa, William Noble, Jian Qiu...